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Three-Toed Sloth as Putative Reservoir of Coxiella burnetii, Cayenne, French Guiana

To the Editor: Q fever is an emerging zoonosis and a major public health concern in French Guiana, a French overseas region located on the northeastern coast of South America (1,2). Most cases occur in the city of Cayenne (3), specifically in the suburbs, where houses are near wooded hills (4). Genotyping performed by using multispacer sequence typing showed that MST17, a unique genotype of C. burnetii, circulates in Cayenne and is responsible for epidemics of Q fever (5). C. burnetii transmission peaks during the rainy season, and the incidence of Q fever usually increases 1–3 months later (6). The animal reservoir of C. burnetii in French Guiana is unknown; previous studies have excluded domestic ruminants, which are known to be C. burnetii reservoirs elsewhere in the world (6). Four serologic surveys showed few C. burnetii–positive opossums, dogs, rodents (Proechimys spp.), bovines, or birds in French Guiana (7). In 2013, using real-time PCR (qPCR) analysis of vaginal swab samples, we showed that 6/158 (3.8%) dogs from Cayenne and 0/206 bats from the coastal area of French Guiana were positive for C. burnetii (Cycle threshold [Ct]<35). One of the positive samples was negative for C. burnetii by qPCR. In addition, serologic tests for C. burnetii were negative for samples from all 37 small ruminants maintained near the outbreak area.

In January 2014, a dead (accidental death) female 3-toed sloth (Bradypus tridactylus) (Figure, panel A) was found on the road near the residence of a Q fever patient. We retrieved the sloth and collected feces, spleen, liver, kidney, lung, and uterus samples and a vaginal swab sample. A total of 16 ticks were removed from the sloth and stored in 70% alcohol.

DNA was extracted from the feces, organs, and ticks by using the BioRobot EZ1 Workstation (QIA-GEN, Courtaboeuf, France). qPCR targeting the repeated insertion sequence IS1111 was performed by using a CFX96 Touch Real-Time PCR Detection System (Bio-Rad, Marne la Coquette, France) as described (8). We confirmed all positive results by performing a second qPCR targeting the IS30α repeated sequence. DNA samples with Ct values ≤35 in both assays were considered positive for C. burnetii. A standard calibration curve quantifying the target IS1111 was generated by using 10-fold serial dilutions of C. burnetii Nine Mile strain. The number of IS1111 intergenic sequences found in the genome of strain C. burnetii MST17 was identical to that for the Nine Mile strain (F. D’Amato, unpub. data); thus, the qPCR that we used was valid for quantifying the number of C. burnetii MST17 IS1111 copies/mL in samples we collected (5).

qPCR analysis showed that the feces were highly positive for C. burnetii; the sample had a low Ct value of 23, corresponding to 7 log_{10} DNA copies/mL (9). The spleen was also positive for C. burnetii; the Ct value was 34, corresponding to 3.6 log_{10} DNA copies/mL. Results for the other samples were negative.

Using morphologic criteria, we identified all 16 ticks collected from the sloth as Amblyomma geayi (Figure, A). A male tick (Amblyomma geayi) found on the 3-toed sloth in this study (photograph by J.M. Berenger). A color version of this figure is available online (http://wwwnc.cdc.gov/eid/article/20/10/14-0694-F1.htm).
Feces from the sloths were highly infectious for C. burnetii. Because sloths live in tall trees and can shed this bacterium in their feces, human contamination might occur through inhalation of infectious aerosols from feces. The high prevalence of C. burnetii infection in ticks also suggests possible transmission through tick bites or from aerosols of tick feces that have been deposited on the skin of animal hosts; such feces can be extremely rich in bacteria and highly infectious (10).

In this 2013 outbreak of Q fever, epidemiologic studies led to the identification of 3-toed sloths as a putative source of C. burnetii infection. Further investigations are needed to confirm the role of sloths as a reservoir for C. burnetii in French Guiana and to implement efficient measures to prevent transmission to humans.

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Marburgvirus Resurgence in Kitaka Mine Bat Population after Extermination Attempts, Uganda

To the Editor: Marburg virus (MARV) and Ravn virus (RAVV), collectively called marburgviruses, cause Marburg hemorrhagic fever (MHF) in humans. In July 2007, 4 cases of MHF (1 fatal) occurred in miners at Kitaka Mine in southern Uganda. Later, MHF occurred in 2 tourists who visited Python Cave, ~50 km from Kitaka Mine. One of the tourists was from the United States (December 2011).