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Laura Fancello, Sébatien Trape, Catherine Robert, Mickaël Boyer, Nikolay Popgeorgiev, et al.. Viruses in the desert: a metagenomic survey of viral communities in four perennial ponds of the Mauritanian Sahara. The International Society of Microbiologial Ecology Journal, 2013, 7 (2), pp.359 - 369. 10.1038/ismej.2012.101 . hal-01785437

HAL Id: hal-01785437
https://amu.hal.science/hal-01785437

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The ISME Journal (2013) 7, 359–369
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ORIGINAL ARTICLE

Viruses in the desert: a metagenomic survey of viral communities in four perennial ponds of the Mauritanian Sahara

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Here, we present the first metagenomic study of viral communities from four perennial ponds (gueltas) located in the central Sahara (Mauritania). Three of the four gueltas (Ilij, Molomhar and Hamdoun) are located at the source of three different wadis belonging to the same hydrologic basin, whereas the fourth (El Berbera) belongs to a different basin. Overall, sequences belonging to tailed bacteriophages were the most abundant in all four metagenomes although electron microscopy and sequencing confirmed the presence of other viral groups, such as large DNA viruses. We observed a decrease in the local viral biodiversity in El Berbera, a guelta with sustained human activities, compared with the pristine Ilij and Molomhar, and sequences related to viruses infecting crop pests were also detected as a probable consequence of the agricultural use of the soil. However, the structure of the El Berbera viral community shared the common global characteristics of the pristine gueltas, that is, it was dominated by *Myoviridae* and, more particularly, by virulent phages infecting photosynthetic cyanobacteria, such as *Prochlorococcus* and *Synechococcus* spp. In contrast, the Hamdoun viral community was characterized by a larger proportion of phages with the potential for a temperate lifestyle and by dominant species related to phages infecting heterotrophic bacteria commonly found in terrestrial environments. We hypothesized that the differences observed in the structural and functional composition of the Hamdoun viral community resulted from the critically low water level experienced by the guelta.

The ISME Journal (2013) **7,** 359–369; doi:10.1038/ismej.2012.101; published online 4 October 2012
Subject Category: microbial ecology and functional diversity of natural habitats
Keywords: viral metagenomics; giant virus; Sahara desert; perennial water pond; Mauritania; Adrar plateau

Introduction

Viruses can colonize virtually all ecosystems on Earth and are found wherever cellular life exists (Le Romancer *et al.*, 2007). In the ocean, viruses (the majority of which are bacteriophages) represent the most abundant biological component of the ecosystem and influence horizontal gene transfer, microbial diversity and biogeochemical cycling (Fuhrman, 1999; Suttle, 2005, 2007). Metagenomics (the sequence-based analysis of the collective genomes contained in an environmental sample) was first applied to environmental viral communities in marine waters 10 years ago (Breitbart *et al.*, 2002). This study demonstrated that the viral fraction represents a vast reservoir of unexplored biodiversity. Since then, viral diversity has been investigated using metagenomics in a wide range of environments, including marine waters (Angly *et al.*, 2006; Culley *et al.*, 2006), freshwaters (Dinsdale *et al.*, 2008a; Djikeng *et al.*, 2009), hot springs (Schoenfeld *et al.*, 2008), soils (Williamson *et al.*, 2005; Fierer *et al.*, 2007), stromatolites and thrombolites (Desnues *et al.*, 2008), and animal-associated biomes (Breitbart *et al.*, 2003; Zhang *et al.*, 2006; Vega Thurber *et al.*, 2008; Ng *et al.*, 2011).

To date, studies on viral diversity have mainly focused on marine waters from temperate regions, and data exploring the extent of viral diversity and ubiquity in arid regions are largely scarce. Unfavorable conditions found in cold or hot deserts limit the development and the activity of eukaryotic life. In such environments, the study of viral assemblages is of particular interest because microbial communities are mainly regulated by viral lysis (Weinbauer, 2004; Laybourn-Parry, 2009). A recent

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Received 10 February 2012; revised 18 June 2012; accepted 16 July 2012; published online 4 October 2012

study in the cold desert of Antarctica used a 0.45-μm size selective metagenomic analysis to show that the viral community of an ice-covered lake presented an unexpectedly high genetic richness, distinct from that of other aquatic viral metagenomes, and was dominated by small single-stranded DNA viruses infecting eukaryotes in the spring and by large double-stranded DNA (dsDNA) viruses (mostly Phycodnaviruses and Mimiviruses) and dsDNA bacteriophages in summer (Lopez-Bueno *et al.*, 2009).

The Sahara is the largest non-polar desert on Earth. In the early Holocene period, the Sahara experienced humid episodes (Kuper and Kröpelin, 2006) that sustained the development of numerous lakes and wetlands; remnants of these aquatic environments still persist today. The Mauritanian Adrar is one of the mountainous massifs of the central Sahara and contains >20 perennial and semi-perennial freshwater bodies. Among them, rocky pools (gueltas) are found in the higher reaches of gorge-like watercourses and are alimented by subterranean seep and rainfall. Some of these gueltas are sites of permanent human settlements and are used for agricultural and animal-farming purposes.

Previous electron microscopy and pulse-field gel electrophoresis studies from bacteriophage particles induced from Namib and Sahara sands have revealed the presence of different morphotypes with genome sizes varying from 45 to 350 kb, suggesting the existence of large viral particles (Prigent *et al.*, 2005; Prestel *et al.*, 2008). The objective of this study was to fill the gaps in our knowledge of the ubiquity and diversity of viruses in hot desert environments. Using metagenomic approaches, we investigated the composition, taxonomy and functional diversity of the viral communities from four gueltas located in the Mauritanian Sahara.

Materials and methods

Geographic location of the sampling sites

During the dry season of 2009 (June), water samples were collected from four different gueltas: Ilij (20°38′046 N, 13°08′490 W), Molomhar (20°35′229 N, 13°08′794 W), Hamdoun (20°19′380 N, 13°08′550 W) and El Berbera (19°59′181 N, 12°49′3744 W) located in the Adrar plateau of Mauritania (Figure 1). Ilij, Molomhar and Hamdoun belong to the same hydrographic network (the Seguellîl wadi basin), whereas El Berbera belongs to a different network (the Timinit wadi basin). The local human population occasionally and permanently occupies the Hamdoun and El Berbera gueltas, respectively. Hamdoun is located close to the main Atar-Nouakchott road and may serve as a temporary open well, whereas El Berbera hosts a permanent human settlement and is a site of intensive date fruit production. During the driest periods of the year, the residual water volume of the Hamdoum guelta can drop to $<2\,m^3$, whereas the volumes of the other gueltas remain between 200 and 500 m^3.

Sampling procedure, virus purification, transmission electron microscopy, nucleic acid extraction and sequencing

During a 2-day mission, one liter of water was collected from each guelta and filtered through a 0.45-μm pore filter. Virus-size particles contained in the filtrate were precipitated on site using PEG (10%)

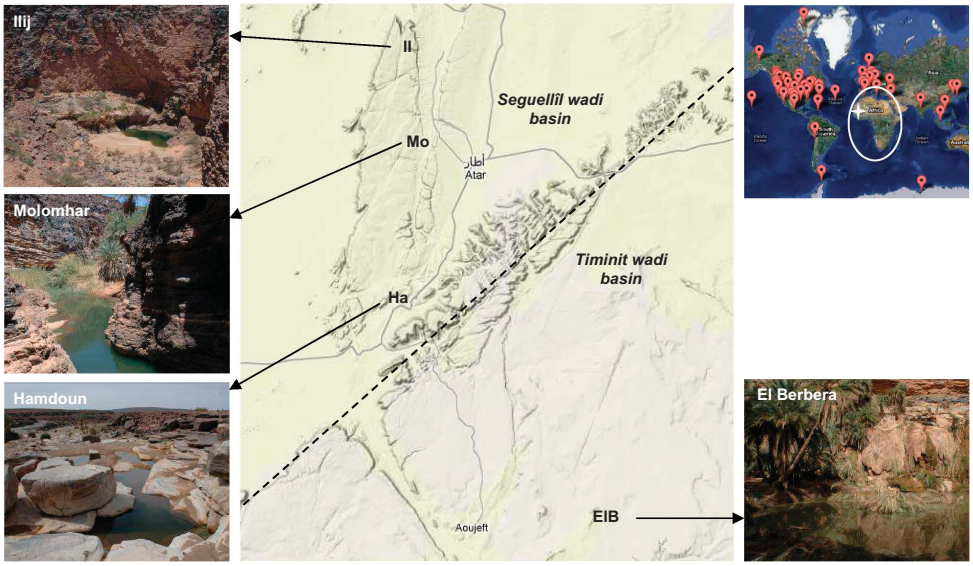


Figure 1 Geographic localization and pictures of the samp ling sites. Central: A Google Earth map of the Adrar region showing mountains (gray) and sandstones (yellow). Il: Ilij guelta, Mo: Molomhar guelta, Ha: Hamdoun guelta, ElB: El Berbera guelta. The dashed line on the map indicates the separation between the two hydrologic systems: the Seguellîl wadi basin and the Timinit wadi basin. Images of the sampling sites are provided on the left and on the right of the map. Upper right: Localization of the 205 environmental metagenomic projects recorded in the Genome On Line Database (GOLD) as of 2012-01-10. Africa is outlined by a circle, and the Adrar plateau of Mauritania is indicated by a star.

and NaCl (1 M final) in bottles maintained at 4 °C in a cool-box. Samples (kept at 4 °C) were brought to the laboratory immediately (within the 48 h) for further processing. Precipitated viral particles were purified using CsCl density gradient ultracentrifugation and DNAse treated as previously described (Thurber *et al.*, 2009). Purified viral particles were stained with 3.5% uranyl acetate and lead citrate and then examined by transmission electron microscopy (TEM) (Philips Morgagni 268D, FEI Co., Eindhoven, The Netherlands). Nucleic acids were extracted using the formamide procedure (Thurber *et al.*, 2009) and amplified using the illustra GenomiPhi V2 DNA Amplification Kit (GE Healthcare Life Sciences, Freiburg, Germany). Because phi29 DNA polymerase has been shown to preferentially amplify circular DNA and genomes from single-stranded DNA viruses (Kim *et al.*, 2008; Kim and Bae, 2011), duplicate reactions were performed to minimize this bias, as previously suggested (Thurber *et al.*, 2009). Amplification products were then pooled, ethanol purified and pyrosequenced on a Roche Applied Sciences (454 Life Sciences, Basel, Switzerland) GS20 platform. Metagenomes are freely accessible on the MG-RAST annotation server with the following accession numbers: El Berbera 2 (4446033.3), Molomhar Guelta (4445718.3), Ilij Guelta (4445716.3) and Hamdoun Guelta (4445715.3).

Taxonomic and functional annotations
Metagenomes were annotated using MG-RAST version 2 (Meyer *et al.*, 2008) with an *E*-value cutoff of 10^{-5}. The MG-RAST server produces automated taxonomic assignments using Blastx searches against the SEED non-redundant database and other accessory databases (rRNA, chloroplast and mitochondrial databases) and also produces metabolic profiles of metagenomes by Blastx comparisons using the SEED-Subsystem data set. Pairwise comparisons of the metabolic profiles were performed using XIPE-TOTEC (Rodriguez-Brito *et al.*, 2006), a non-parametric pairwise bootstrap statistical test that was specifically developed for metagenomic functional comparisons and is based on median difference analysis. This test locates statistically significant differences and identifies subsystems that are overrepresented in each comparison. The confidence level chosen for the test was 98%.

Sequence analysis
The GC content of the four metagenomes was analyzed using the geecee function of EMBOSS. The average GC fraction was computed for each metagenome as a whole or separately for subsets of bacterial- and viral-annotated reads.

Assembly and phylogeny
The assembly of each metagenome was performed using the Genome Sequencer (GS) *De Novo* Assembler version 2.0.01 (Roche Diagnostics, Meylan, France), an application especially suited to the analysis of GS-FLX data. We chose a minimum overlap length of 20 bp and a minimum overlap identity of 95%. We only kept contigs longer than 300 bp for subsequent analyses because the average read length was 251–258 bp. Open Reading Frames (ORFs) were searched on large contigs (>1500 bp) by Prodigal (Hyatt *et al.*, 2010) and MetaGeneMark (Zhu *et al.*, 2010). Phylogenetic trees were constructed for ORFs with at least 10 homologs, according to a Blastx search against the NCBI non-redundant database (*E*-value $<1e^{-10}$). An ORF and its homologs were aligned using MUSCLE (Edgar, 2004), and the alignment was curated using Gblocks (Castresana, 2000). Phylogenetic trees were constructed using PhyML (Guindon *et al.*, 2010), with 100 bootstrap replicates, and visualized using MEGA v5 software (Tamura *et al.*, 2011). A specific research of phages and prophages has also been performed on assembled contigs by a Blastn search (*E*-value $<1e^{-05}$) against the Aclame database (Leplae *et al.*, 2004).

Mapping
For each of the most abundant organisms found in the MG-RAST analysis, metagenomic reads were mapped (that is, aligned against a reference sequence) on the genome of that organism. Mapping was performed using the GS Reference Mapper version 2.0.01 (Roche).

Population modeling
Information about community structure and diversity was obtained for each metagenome using the following workflow: (i) computation of the community contig spectrum using the free Circonspect software, (ii) evaluation of average genome sizes using GAAS free software (Angly *et al.*, 2009) with an *E*-value cutoff of 10^{-3} and (iii) mathematical modeling of the community structure and diversity by PHACCS (Angly *et al.*, 2005).

Phylogenetic tests
Several phylogenetic tests were performed using the FastUniFrac tool (Hamady *et al.*, 2009) to find statistically significant differences among the four metagenomes. The analyses were performed on the subset of viral sequences of each metagenome. For each metagenome, FastUniFrac uses phylogenetic information to assemble metagenomic sequences into a tree. P-test and UniFrac metric capture significant diversity between the trees associated with the different metagenomes, and they account only for tree topology or for both tree topology and branch length, respectively. The two tests can be used for multiple or pairwise comparisons or to compare one particular tree with all others.

Principal Component Analysis (PCA) and hierarchical clustering were also performed using FastUniFrac. The robustness of the clustering results to the sampling effort and evenness was determined using the Jackknife Environment Clusters analysis option.

Comparative metagenomics of viral communities from freshwater environments

A multiple comparison of the phylogenetic profiles of different natural and non-natural freshwater viral communities was performed using the MG-RAST server (Meyer *et al.*, 2008). Ten viral metagenomes were compared; the four analyzed in this work, two from two different temperate freshwater lakes (Roux *et al.,* 2012), one from an Antarctic lake sampled in the spring and summer seasons (Lopez-Bueno *et al.* 2009), and two from an aquaculture system (Dinsdale *et al.*, 2008a). The phylogenetic profile was based on the sequence taxonomic assignment according to a Blastx search (*E*-value $< 1e^{-05}$) against the NCBI GenBank non-redundant database. Multiple comparisons were performed by PCA on the MG-RAST server using normalized values and a Bray-Curtis distance matrix. *P*-values were computed on the MG-RAST server (Meyer *et al.*, 2008).

Results

Taxonomic composition of the viral metagenomes

A total of 82 814 818 bp of sequence was generated from the four samples (Ilij ~17 Mbp, Molomhar ~25 Mbp, Hamdoun ~15 Mbp, El Berbera ~24 Mbp), corresponding to 324 603 sequences with an average length of 250 bp. Annotation of the sequence fragments by MG-RAST using an *E*-value cutoff of $1e^{-05}$ indicated that 70.50–83.21% of these fragments had no significant hits to known sequences stored in the SEED non-redundant database or other accessory databases (Figure 2). According to the MG-RAST annotation, 8.06–34.42% of the known reads were classified as viruses. The majority of viral reads belonged to dsDNA viruses (Table 1) and, among these, >92% matched with Caudovirales. Sequences belonging to the *Myoviridae* were the most abundant in all metagenomes followed by *Podoviridae* and *Siphoviridae*. The presence of tailed phages was confirmed by TEM (Figures 3a and b). Viral morphotypes were usually between 50 and 200 nm in diameter, but some viral particles with diameters <50 nm (Figures 3d–f, arrows) and >200 nm (Figure 3c) were also observed. Among the dsDNA viruses, sequences belonging to eukaryotic viruses were also found (Tables 1 and 2). Four out of the seven families of nucleocytoplasmic large DNA viruses group were represented (Table 1, in bold). Viruses from the *Phycodnaviridae* (infecting algae) and *Mimiviridae* (infecting amoebas and algae) were more abundant in Hamdoun (4.25%) and El Berbera (3.18%). *Poxviridae* sequences were also more frequently found in the El Berbera metagenome. Only a few reads were associated with single-stranded DNA viruses. The majority of these reads were found in the Molomhar metagenome and were related to *Microviridae* (2.92%). To analyze the taxonomic composition more accurately, we used GAAS that normalizes the number of hits for the genome size and then provides a more realistic description of species abundances. According to the GAAS analysis, the most represented viral genotype was that of *Prochlorococcus* phage, found in three out of the four gueltas (Ilij, Molomhar and El Berbera) with relative abundances ranging between 31.54 and 55.24% (Table 2). In contrast, Hamdoun

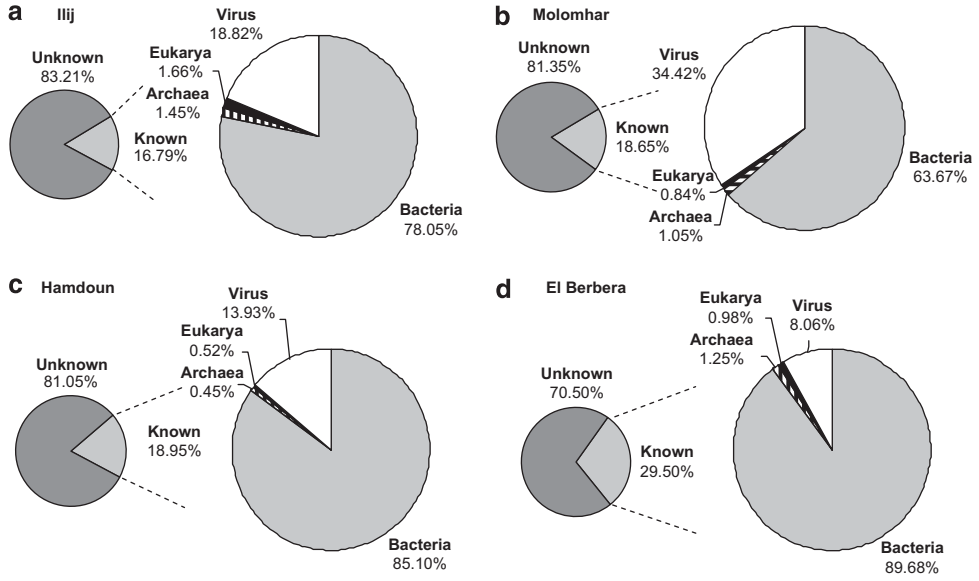


Figure 2 Reads classification according to their Best Blast Hit (*E*-value $< 10^{-5}$) in the MG-RAST analysis. (**a**) Ilij guelta, (**b**) Molomhar guelta, (**c**) Hamdoun guelta and (**d**) El Berbera guelta.

Table 1 Classification of reads hitting viral sequences

Group	*Order*	*Family*	*Ilij (%)*	*Molomhar (%)*	*Hamdoun (%)*	*El Berbera (%)*
		Unclassified	5.78	2.86	5.16	5.92
	Caudovirales	*Myoviridae*	63.33	73.04	48.03	56.43
		Podoviridae	11.75	8.45	22.61	18.92
		Siphoviridae	12.50	8.20	16.39	11.93
	Herpesvirales	*Herpesviridae*	0.05	0.02	0.00	0.00
dsDNA	–	*Tectiviridae*	0.05	0.06	0.00	0.00
	–	***Iridoviridae***	0.67	1.11	0.83	0.54
	–	***Phycodnaviridae***	2.08	1.20	4.25	2.25
	–	***Poxviridae***	0.28	0.02	0.38	0.54
	–	***Mimiviridae***	1.90	0.84	0.99	3.18
	–	*Baculoviridae*	0.00	0.00	0.00	0.15
ssDNA	–	*Circoviridae*	0.00	0.14	0.00	0.05
	–	*Microviridae*	0.00	2.92	0.15	0.09
	–	*Geminiviridae*	0.00	0.02	0.08	0.00
	–	*Nanoviridae*	0.05	0.14	0.08	0.00
	–	*Parvoviridae*	0.00	0.00	0.08	0.00
Retroviridae	–		0.00	0.00	0.08	0.00
Unclassified phages/viruses	–		1.56	0.98	0.91	0.00

Abbreviations: dsDNA, double-stranded DNA; ssDNA, single-stranded DNA.
Assignment was made according to the best Blastx hit (*E*-value $<10^{-5}$) in the MG-RAST analysis.

was dominated by viruses that infect members of the genus *Microbacterium* (*Microbacterium* phage Min1), which represented >44% of the total viral genotype abundance (Table 2).

Although no bacterial cells could be detected via electron microscopy, 63.67–89.68% of the reads were classified as bacterial in the viral metagenomes (Figure 2; Supplementary Table 1). Using GAAS, all bacteria-annotated reads were dominated by sequences related to the *Acinetobacter* genus (Supplementary Table 2). Between 3 and 20 sequences matching bacterial 16S rRNA genes were also found for each metagenome (Supplementary Table 3).

Functional annotation and metabolic analysis

The metabolic profile of the four metagenomes was explored using MG-RAST, which assigns sequences to metabolic categories based on their Best Blastx Hit against the SEED database (*E*-value $<1e^{-05}$). Only 6.22–17.43% of the sequences could be functionally classified in this way. The most represented categories were related to the metabolism of carbohydrates, amino acids, proteins, cofactors, vitamins, DNA, and nucleosides/nucleotides (Figure 4). We compared these data with the metabolic profile derived from the combined analysis of 42 viral metagenomes (subterranean, hypersaline, marine, aquaculture freshwater, coral, microbialites, fish, terrestrial animals and mosquito) described in a previous study (Dinsdale *et al.*, 2008a). The guelta metagenomes were depleted in virulence subsystems compared with the average value found for the other 42 viral metagenomes. Metabolic profile comparisons using XIPE-TOTEC showed that respiration, regulation and cell signaling, and motility and chemotaxis subsystems were overrepresented in the Hamdoun metagenome compared with the other gueltas (*P*-value <0.02). Deeper in the respiration subsystem hierarchical levels, the electron donating reaction of the Hamdoun metagenome was dominated by the respiratory dehydrogenase I subsystem, which was mainly represented by the proline dehydrogenase. In contrast, the other three gueltas were dominated by the NAD(P)H dehydrogenase complex, which is classified as a respiratory complex I subsystem. An overrepresentation of RNA metabolism was also evidenced by the XIPE-TOTEC analyses of the El Berbera metagenome, whereas the Molomhar metagenome displayed a statistically significantly higher number of sequences related to photosynthesis and nucleoside/nucleotide metabolism (*P*-value <0.02).

Assembly, contig analysis and mapping

Contigs were assembled using the GS *De Novo* Assembler, and only contigs longer than 300 bp were kept (Supplementary Table 4). The average contig length was from 742 to 990 bp, and large contigs were also obtained (for example, 55 kbp in El Berbera and 52 kbp in Hamdoun). Overall, 29.90–37.06% of the contigs had similarities to phage and prophage sequences in the Aclame database (Supplementary Table 5). Viral assemblies were dominated by phage genomes. However, mapping to fully sequenced genomes of the most abundant phages in the metagenomes resulted in low coverages (<5%; Supplementary Figure 1). Low coverage was also found for plasmids; the maximum plasmid coverage was 14.24% for the *Acinetobacter venetianus* pAV2 genome.

We were able to assemble 30.33–68.72% of all reads (Supplementary Table 4). Interestingly, between 34.98 and 93.03% of the 'unknown' reads

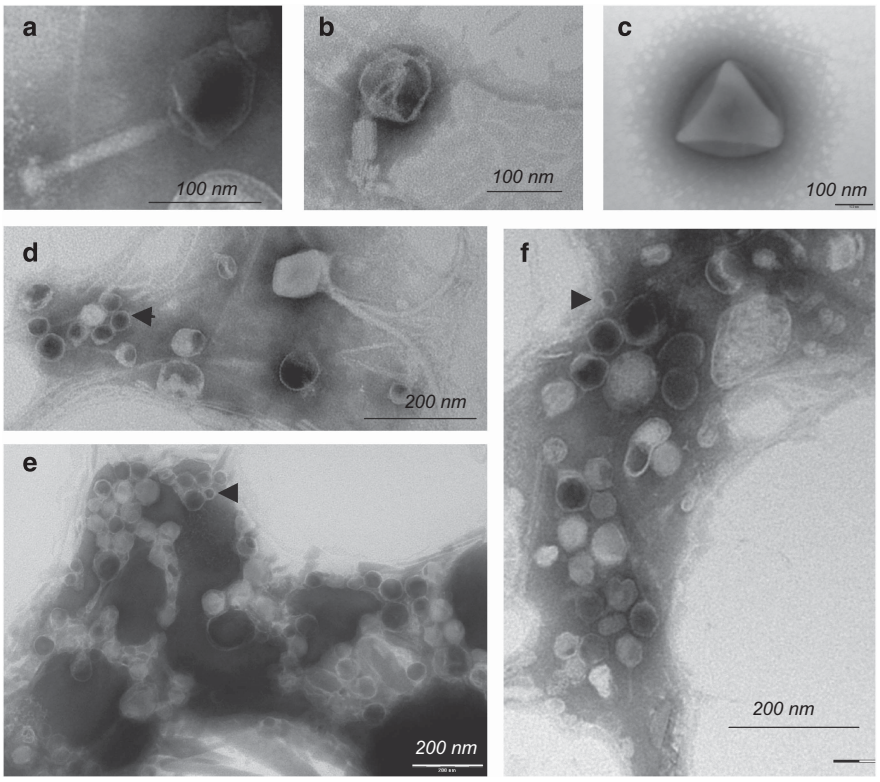


Figure 3 Viral morphotypes observed under TEM in the gueltas of the Adrar plateau. Example of tailed phages belonging to the Myoviridae family (**a**, **b**). Viral morphotypes were usually between 50 and 200 nm in diameter (**d–f**), but some small viral particles with diameters <50 nm (**d–f**, arrows) and large Mimivirus-like particles (**c**) were also observed. Images (**a**) and (**d**) are from Ilij, (**b**) and (**f**) are from Hamdoun, and (**c**) and (**e**) are from Molomhar.

Table 2 Most represented viral genotypes among the viral hits according to GAAS analysis

Metagenome	*Viral species*	*Relative abundance (%)*	*Host*
Ilij	***Prochlorococcus* phages**	**55.2449**	B
	Burkholderia phages	18.9451	B
	***Synechococcus*phages**	**15.9518**	B
	Roseobacter phage SIO1	5.8638	B
	Acanthocystis turfacea Chlorella virus 1	2.4366	E
	Aeromonas phages	1.3597	B
Molomhar	***Prochlorococcus* phages**	**46.0733**	B
	***Synechococcus* phages**	**35.6391**	B
	Mycobacterium phages	10.2428	B
	Bordetella phages	2.0430	B
	Acyrthosiphon pisum secondary endosymbiont phage	1.1061	E
	Acanthocystis turfacea Chlorella virus 1	1.0322	E
Hamdoun	*Microbacterium* phage Min1	44.3371	B
	***Synechococcus* phages**	**29.9623**	B
	***Prochlorococcus* phages**	**11.5155**	B
	Acanthocystis turfacea Chlorella virus 1	10.7050	E
	Acanthamoeba polyphaga mimivirus	3.4801	E
El Berbera	***Prochlorococcus* phages**	**31.5358**	B
	***Synechococcus* phages**	**26.8707**	B
	Mycobacterium phages	14.3964	B
	Lactobacillus phage phiJL-1	9.7423	B
	Burkholderia phage phi644-2	7.3403	B
	Spodoptera litura NPV	2.7741	E
	Musca domestica salivary gland hypertrophy virus	2.6775	E
	Acanthocystis turfacea Chlorella virus 1	2.4808	E
	Ostreococcus virus OsV5	1.7077	E

Abbreviations: B, Bacteria; E, Eukaryotes.
Only viral genotypes with a relative abundance superior to 1% are indicated. Viral species infecting cyanobacteria are shown in bold.

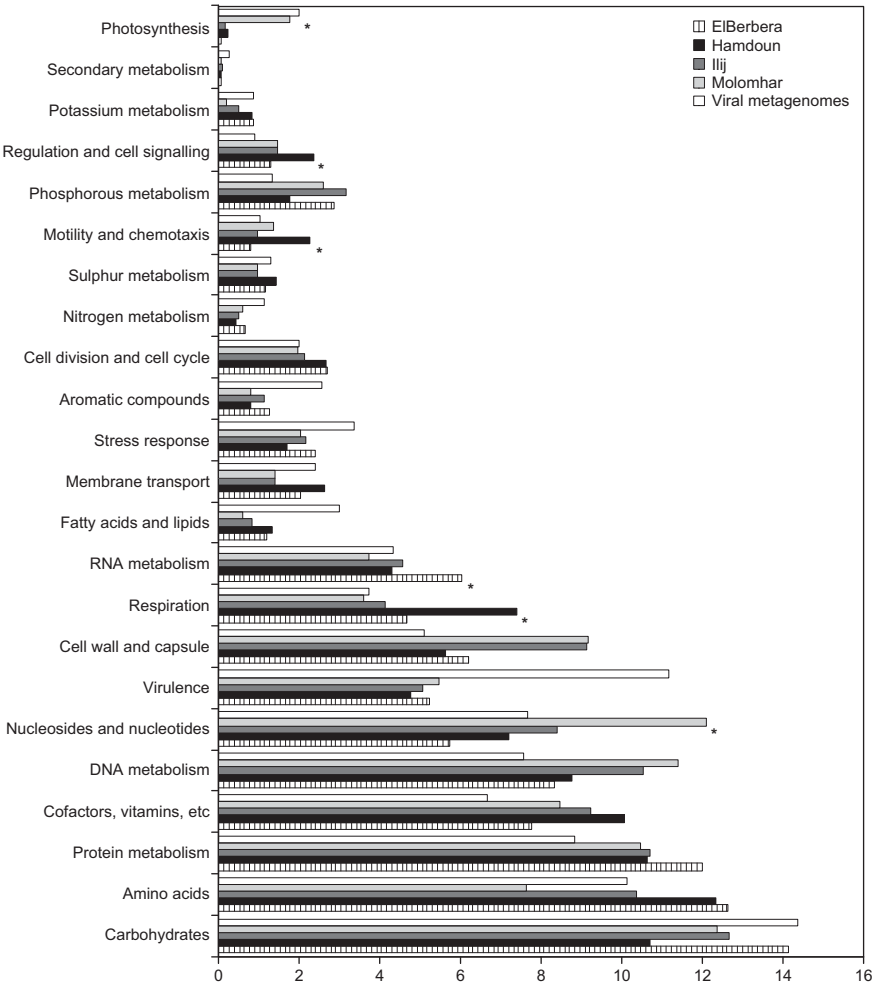


Figure 4 Relative abundances of sequences assigned to each metabolic subsystem by MG-RAST. The metabolic categorization is based on the sequences best Blast hits in the SEED database curated subsystems (*E*-value $<1e^{-05}$). Asterisks: metabolic subsystems for which pairwise comparisons were performed by XIPE-TOTEC to identify statistically significant differences ($P<0.05$) between the four guelta metagenomes.

identified by the MG-RAST annotation system were assembled into contigs (Supplementary Table 4). The assembly of these unknown reads into contigs could facilitate their taxonomic assignment. Indeed, short sequences are less likely than long sequences to retrieve statistically significant similarities in Blast searches and sequence assembly into longer contigs is helpful to overcome this difficulty (Wommack *et al.*, 2008). Moreover, long contigs can contain unknown reads and reads with far homologies to known sequences, which are suggestive of the putative phylogenetic origin of the whole contig. The largest contigs (>1500 bp) were annotated by ORF prediction and Blast search (Supplementary Table 6). When possible, phylogenetic analysis was performed to confirm the origin of the predicted ORFs (Supplementary Figure 2). A few large contigs contained a relevant proportion of predicted ORFs with similarities to phage sequences and coding for some specific conserved phage proteins, that is, terminases, structural proteins (mainly related to Caudovirales tail structures) and phage DNA polymerases (Supplementary Table 6). It has been previously shown that viral genomes contain more ORFans (that is, ORFs without homologs in the databases) than do bacteria (~30 and ~10% for viral and bacterial genomes, respectively) (Yin and Fischer, 2006; Boyer *et al.*, 2010) and that viral (meta)genomes tend to be more AT rich than those of their hosts (Rocha and Danchin, 2002; Willner *et al.*, 2009). Thus, contigs with >50% ORFans, low GC%, and for which conserved viral protein-encoding genes have been identified, can confidently be considered as of viral origin (for example, contigs ElBerbera_882 or Hamdoun_439 in Supplementary Table 6).

Community phylogenetic structure and diversity across sampling sites

Viral community structure and diversity estimations were performed using the PHACCS analysis system

on each metagenome (Supplementary Figure 3). Briefly, contig spectra were generated using the Circonspect tool and the average genome size was estimated by GAAS. These parameters were passed to PHACCS for alpha-diversity analysis. Computed community structures (defined by richness (R), evenness (E) and diversity (H')) are graphically represented as rank-abundance curves in Supplementary Figure 3. Based on the obtained results, the samples with the highest viral diversity index were the pristine Ilij and Molomhar gueltas (H' = 4.83 and H' = 4.33, respectively), followed by El Berbera (H' = 4.19) and Hamdoun (H' = 2.21). The phylogenetic composition of the viral communities of the four metagenomes was then considered, and comparisons were computed using the FastUniFrac tool on the subset of viral annotated metagenomic sequences. Statistically significant differences were measured between two samples (P-value < 0.05) using the 'UniFrac significance analysis' test and further confirmed using the P-test. PCA, which was used to visualize multiple comparisons between samples (Supplementary Figure 4), showed that Hamdoun is isolated from the other samples. To evaluate the robustness of this clustering pattern, we performed a Jackknife environment cluster analysis. The results of this bootstrap procedure confirmed the confidence in the Hamdoun cluster node.

Comparisons with viral communities from other freshwater environments

The phylogenetic profile of the four gueltas was compared with that of other natural or non-natural freshwater environments, two temperate freshwater lakes (Roux *et al.*, 2012), an Antarctic lake sampled in the spring and summer seasons (Lopez-Bueno *et al.*, 2009) and two freshwater samples from a human-controlled aquaculture system (Dinsdale *et al.*, 2008a; Figure 5). As a phylogenetic profile representation, we used the metagenomic sequences classification according to their best Blast hit in a Blastx search against the NCBI GenBank non-redundant database (E-value < $1e^{-05}$). Multiple comparisons were performed and visualized by PCA on the MG-RAST server. The results showed a geographic clustering pattern with the Mauritanian gueltas clustering together, separate from the others (and Hamdoun again separated from the other three Mauritanian gueltas) (Figure 5). Similarly, metagenomes from temperate natural and artificial freshwaters group together and are separate from the metagenomes of other environments. The two metagenomes from the Antarctic lake did not group together, which is consistent with the phylogenetic profile differences observed between the spring and summer communities from this lake (Lopez-Bueno *et al.*, 2009). The metagenome-clustering pattern showed statistically significant differences in the viral domain (P-value = 0.006).

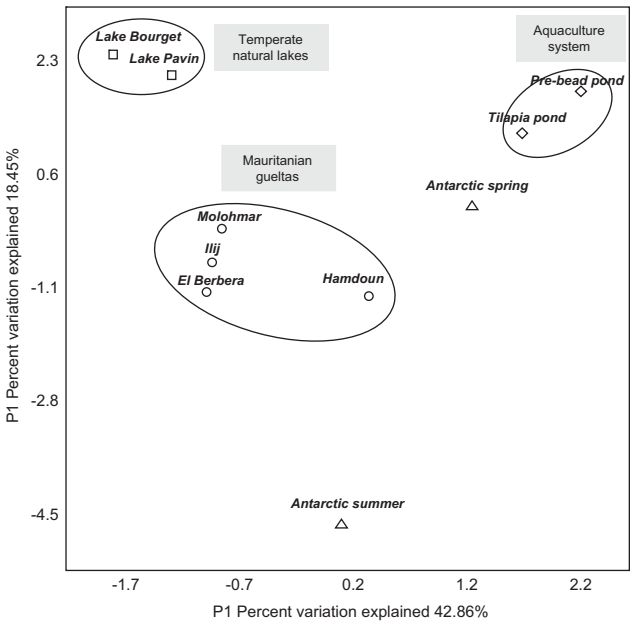


Figure 5 First two principal coordinates from the principal coordinate analysis of the viral communities in freshwater samples from different environments. The PCA was run in MG-RAST to visualize the overall patterns of variation between the samples.

Discussion

With the advent of metagenomics, an increasing number of studies describing viral and bacterial diversity have been conducted. Currently, only a few investigations have focused on viral assemblages in freshwaters, and most of them concern freshwaters from non-natural or polluted ecosystems. For example, viral communities have been described from aquaculture ponds (Dinsdale *et al.*, 2008a; Rodriguez-Brito *et al.*, 2010), a cattle farm pond (Rooks *et al.*), reclaimed and potable waters (Rosario *et al.*, 2009), hydrocarbon-polluted groundwater (Abbai *et al.*, 2012), and a man-made recreational lake in MD, USA (Bench *et al.*, 2007). Viral communities in natural freshwater systems have only been described in an ice-covered lake in Antarctica (Lopez-Bueno *et al.*, 2009) and, more recently from two temperate freshwater lakes in France (Roux *et al.*, 2012). By combining electron microscopy and metagenomics, we provide here the first comprehensive analysis of viral communities from freshwater ponds in the Sahara desert of Mauritania.

Most studies focusing on viral diversity in the environment use a 0.2-μm filtration step to separate viruses and bacteria on size criteria. One drawback of this method is that it may fail to recover large viral particles, which are supposed to be common in aquatic ecosystems (Claverie, 2005). However, using a 0.45-micron pore size filter, Lopez-Bueno *et al.* (2009) were able to identify sequences associated with large dsDNA viruses (mainly from the

Phycodnaviridae and *Mimiviridae* families) from an Antarctic freshwater lake in summer. In this study, up to 6.51% of the sequences matched large dsDNA viruses, and the presence of large viral particles (>200 nm) with Mimivirus-like morphologies was confirmed by electron microscopy (Figure 3c). These results further support that large DNA viruses are common in the environment (Ghedin and Claverie, 2005; Monier *et al.*, 2008a, b) and that the 0.2-μm filtration step currently used to prepare environmental viral metagenomes most likely leads to an underestimation of their genetic diversity.

No bacterial cells were observed under electron microscopy, and the number of reads annotated as bacteria (Figure 2) was similar to those reported for other environmental viral metagenomes (Edwards and Rohwer, 2005), indicating a low bacterial contamination of the metagenomes. In addition, the high proportion of unassigned sequences and relatively low number of 16S rRNA matching sequences (Supplementary Table 3) supported a viral origin for the bacterially annotated reads. Because bacterial genes can be packaged into generalized transducing phage particles (Beumer and Robinson, 2005; Ghosh *et al.*, 2008; Del Casale *et al.*, 2011), the bacterial-like sequences in the guelta metagenomes might come from excised prophages mistakenly annotated as bacterial and/or from genes of bacterial origins that were transferred to their phages.

Blast searches performed on the viral metagenomes showed that >70% of the sequences before assembly did not have homologs in current sequence databases (Figure 2). This result is consistent with results of previously published viral metagenomic projects (Breitbart *et al.*, 2002; Desnues *et al.*, 2008; Lopez-Bueno *et al.*, 2009) and again emphasizes that most of the biological diversity in the viral world is still unknown. In this case, sequence assembly using low stringency parameters (20 bp coverage and 95% identity) was of particular interest in classifying the unknown sequences. For example, the Hamdoun metagenome contained >90% unknown reads that could be assembled into contigs (Supplementary Table 4). The downstream identification of structurally conserved viral genes in these contigs (Supplementary Table 6) has provided information about the putative viral origin of these ORFans.

The guelta viral metagenomes were largely dominated by Caudovirales reads, and TEM confirmed the presence of tailed phages (Figure 3) along with other viral morphotypes. Caudovirales are common in the environment and are the dominant viral type recovered from metagenomic analyses in marine environments (Breitbart *et al.*, 2002; Suttle, 2005). Myoviruses, Siphoviruses and Podoviruses were also the most frequently observed viral particles in samples of Namib and Sahara desert sands after the mitomycin C induction of prophages and sonication to release pseudo-lysogens (Prigent *et al.*, 2005; Prestel *et al.*, 2008). The Molomhar metagenome presented the largest number of reads (73% of all reads) that were related to Myoviruses and only 8.2% of reads represented Siphoviruses. At a deeper taxonomic level, viruses infecting photosynthetic bacteria (for example, *Prochlorococcus* and *Synechococcus* phages; in bold, Table 2) were the most abundant in both absolute percentage and rank in the El Berbera, Ilij and Molomhar metagenomes.

Despite being a site for a permanent human settlement, the El Berbera viral community presented a viral community structure similar to those of the Ilij and Molomhar pristine gueltas. It has been previously shown that human activities can affect the diversity and the composition of microbial communities and, thus, their viral predators (reviewed in Horner-Devine *et al.*, 2004). For instance, microbial and viral communities from four coral atolls in the Pacific Ocean dramatically changed along a gradient of human disturbance; the most human-impacted atoll was dominated by heterotrophic microbes, including a large percentage of potential human pathogens (Dinsdale *et al.*, 2008b). In this study, the index of viral biodiversity was inversely correlated with human presence (Supplementary Figure 3), with El Berbera displaying a lower diversity index than the pristine Ilij and Molomhar gueltas. In addition, reads matching *Spodoptera litura* NPV, a baculovirus infecting *S. litura* (Lepidoptera: *Noctuidae*) a crop pest in tropical regions (Rao *et al.,* 1993), were found in the El Berbera metagenome; its presence is most likely linked to the agricultural activity (date production) that developed around the guelta. However, no significant change in the taxonomic structure of the viral communities was observed between the El Berbera human-populated and the Ilij and Molomhar non-populated gueltas. This stability reflects either that the magnitude of human disturbance is weak enough to be absorbed by the system or that the sequencing depth is not sufficient to statistically support finer differences in the phylogenetic profiles between the El Berbera, Ilij and Molomhar metagenomes.

In comparison with the Ilij, Molomhar and El Berbera metagenomes, the Hamdoun metagenome contains dramatically fewer reads matching Myoviruses but more reads related to Siphoviruses (Table 1). This result shows that as it is the case for terrestrial and sediment phage communities (Breitbart *et al.*, 2004; Williamson *et al.*, 2007), viruses with the potential for temperate lifestyles were common in this environment. This is also confirmed by the viral community taxonomic profile, which is dominated by the *Microbacterium* phage Min1 (Akimkina *et al.*, 2007) a potentially temperate phage that infects *Microbacterium* sp., a versatile heterotrophic bacteria that is frequently isolated from the rhizosphere and soils (Takeuchi and Hatano, 1998). The presence of viruses with the ability for being temperate in the free-viral fraction may be related to high nutrient availability, high bacterial density or to environmental stress that

leads to virus induction (McDaniel *et al*., 2002; Breitbart *et al*., 2004). For decades, the Sahara has experienced a dramatic rainfall deficit, and a recent study has stressed that, with only 1.7 m^3 of remaining water in July 2007, Hamdoun is one of the most endangered gueltas of the Adrar plateau (Trape, 2009). We hypothesize that the high proportion of induced lysogens in the Hamdoun viral community compared with the other gueltas reflects stress associated with the unprecedentedly low water content of the pond. Further studies will be required (for example, after the rainy season) to confirm this hypothesis and to determine whether the current structure of the viral community is maintained over time.

Acknowledgements

We thank Florent Angly for his help with GAAS and Jean-François Trape for the valuable assistance during the field surveys. This work was funded by the Centre National de la Recherche Scientifique (crédits récurrents).

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