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Selective On/Off-Nitroxides as Radical Probes to Investigate Non-radical Enzymatic Activity by Electron Paramagnetic Resonance

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Abstract: A nitroxide carrying a peptide specific to the binding pocket of the serine proteases chymotrypsin and cathepsin G is prepared. This peptide is attached as an enol ester to the nitroxide. Upon enzymatic hydrolysis of the peptide, the enol ester moiety is transformed into a ketone moiety. This transformation affords a difference of 5G in phosphorus hyperfine coupling constant between the electronic paramagnetic resonance (EPR) signals of each nitroxide. This property is used to monitor the enzymatic activity of chymotrypsin and cathepsin G by EPR. Michaelis constants were determined and match those reported for conventional optical probes.

In normal tissues enzymatic proteolytic activities are tightly regulated spatially and temporally. However, numerous diseases such as pulmonary inflammatory diseases—asthma, cystic fibrosis (CF), chronic obstructive pulmonary disorder (COPD; e.g., emphysema), acute respiratory distress syndrome and α -1-antitrypsin deficiency—as well as cancers, arthritis and pancreatitis are concomitant to unusually persistent enzymatic ac-

tivities. COPD alone ranks as the fifth leading cause of mortality worldwide which affects an estimated 175 million people and accounts for 3.2 million deaths in 2015.^[1] Therefore, noninvasive or slightly invasive diagnostic/monitoring techniques are of the utmost interest for efficient management of patients. Molecular imaging of the proteolytic activity is a very appealing approach and has been mainly applied using internally quenched fluorescent substrates. Despite suitable enzymatic constants^[2] this method suffers from several drawbacks: incomplete quenching of substrate fluorescence, limited tissue penetration of light, difficult skull imaging or imaging in large animals and three-dimensional images require reconstruction. Due to superior true 3D coding and the used wavelengths interfering weakly with tissues, magnetic resonance imaging methods are the most attractive approach to circumvent the limits of fluorescent techniques. Free radicals such as nitroxides or trityl radicals are stable enough in physiological conditions (in vitro and in vivo) to be detected by electronic paramagnetic resonance (EPR).^[3] Interestingly, EPR is a sensitive technique for the detection of free radical, suitable in visible light-opaque media and which reports only on radical species affording, in general, a rather simple signal. Unfortunately, due to the very fast relaxation of free electron, EPR imaging (EPRI) is at the moment slow and insufficiently resolved.^[4] On the other hand, magnetic resonance imaging (MRI), which is one of the most powerful methods that delivers exquisite anatomic detail, cannot be applied due to its low sensitivity which limits its applicability in molecular imaging of enzymatic activity.^[5] Nevertheless, an emerging double resonance experiment, Overhauser-enhanced magnetic resonance imaging (OMRI), which transfers a part of the higher magnetization of a free radical (nitroxide in this case) to the protons neighboring water molecules (through the electron-proton Overhauser effect), affords a brighter zone on the image and, in turn, the possibility to visualize the site of enzymatic activity.^[6] This technique requires the use of a nitroxide providing an EPR signal sensitive to this activity. Indeed, free organic radicals are currently used for oximetry,^[7,8] redox status imaging,^[9,10] for pH measurement,^[11–14] and for water content measurement.^[15] However, none of these approaches can be employed to investigate nonradical enzymatic activity either by EPR or OMRI. Drescher and colleagues have shown that the signal of a nitroxide might be used to report on enzymatic activity using a very tedious experimental procedure.^[16] Preliminary investigations in our groups^[17–19] indicated that nonradical enzymatic activity can be

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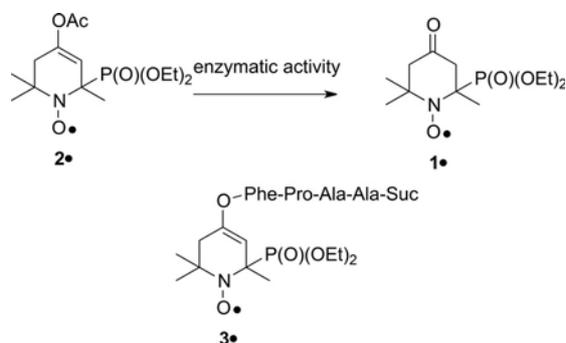
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investigated both by EPR and OMRI using signal changes based either on the linewidth broadening effect^[17,18] or on the shift^[19] of hyperfine coupling constant of a nitroxide. Regardless its lack of selectivity, the approach based on shifting-nitroxides **1**/**2**' (Scheme 1) was very promising and encouraged us



Scheme 1. Enzymatic activity on enol acetate **2**' releasing **1**'.

to develop radical probes exhibiting high selectivity for enzymes. Here we report the synthesis of a nitroxide probe **3**' (Scheme 1) specific to chymotrypsin and cathepsin G. Its EPR signal and sensitivity to enzyme catalysis are reported. In brief, the reported peptide-linked nitroxide is highly selective to these two enzymes and reports activities in nanomolar concentrations of enzymes, best among the array of available conventional nonradical probes.

Since enzymes are known to be substrate specific, the peptide-nitroxide **3**' was designed based on a well-documented substrate for chymotrypsin Suc-Ala-Ala-Pro-Phe.^[20] The substrates Suc-Ala-Ala-Pro-Phe-(*S*)-**2**' ((*S*)-**3**') and Suc-Ala-Ala-Pro-Phe-(*R*)-**2**' ((*R*)-**3**'), which are thought to be selective for the enzymatic activity of chymotrypsin and cathepsin G, were prepared (see the Supporting Information). For this purpose, nitroxide **1**' was prepared in eight steps from commercially available starting materials **1** (Scheme 2A). The first three steps were performed as previously reported to afford **2**.

Then, Kabachnik–Fields reaction using standard conditions (NH_3 , HP(O)(OEt)_2) afforded the amino-phosphonate **3** in 37% yield. Consequently, the reaction was repeated under different conditions (e.g., $\text{Mg}(\text{ClO}_4)_2$, 100°C , ammonium acetate which also failed to improve the yield of the reaction.^[21–25] A two-step procedure (generation of imine followed by addition of diethyl phosphite)^[25] also failed to afford the amino-phosphonate **3** in good yield. Finally, 10 mol % of FeCl_3 was found to be an efficient catalyst for this reaction providing the amino-phosphonate **3** in 87% yield.^[26] Then, **3** was converted into **1**' in five steps as previously reported.^[19] Enantiomers of **1**' were separated by using a Chiralpak-IE column (**1a**', $t_R=6.21$ min, **1b**', $t_R=7.64$ min in 1:1 heptane/ethanol at 1 mL min^{-1} flow rate).

In order to determine the absolute configuration of the enantiomers of **1**', enantiomer **1a**' ($t_R=6.21$ min in 1:1 heptane/ethanol) was converted into alkoxyamine **4a** and **4b**^[27] for which diastereoisomers were easily purified by silica gel chromatography (Scheme 2B). The less polar ketone **4a** was reduced in the presence of NaBH_4 , and then esterified using

(1*S*,4*R*)-(-)-camphanic chloride^[28] as the reference chiral center affording 55:45 mixture of diastereoisomers **5a** and **5b**, respectively (Scheme 2B and the Supporting Information). After purification, crystals were grown for X-ray crystallography.^[29] The XRD structure displays *R* configuration for the chiral center in nitroxide **1a**' (Figure 1 and Supporting Information).

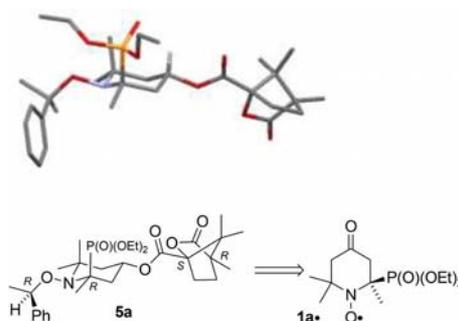
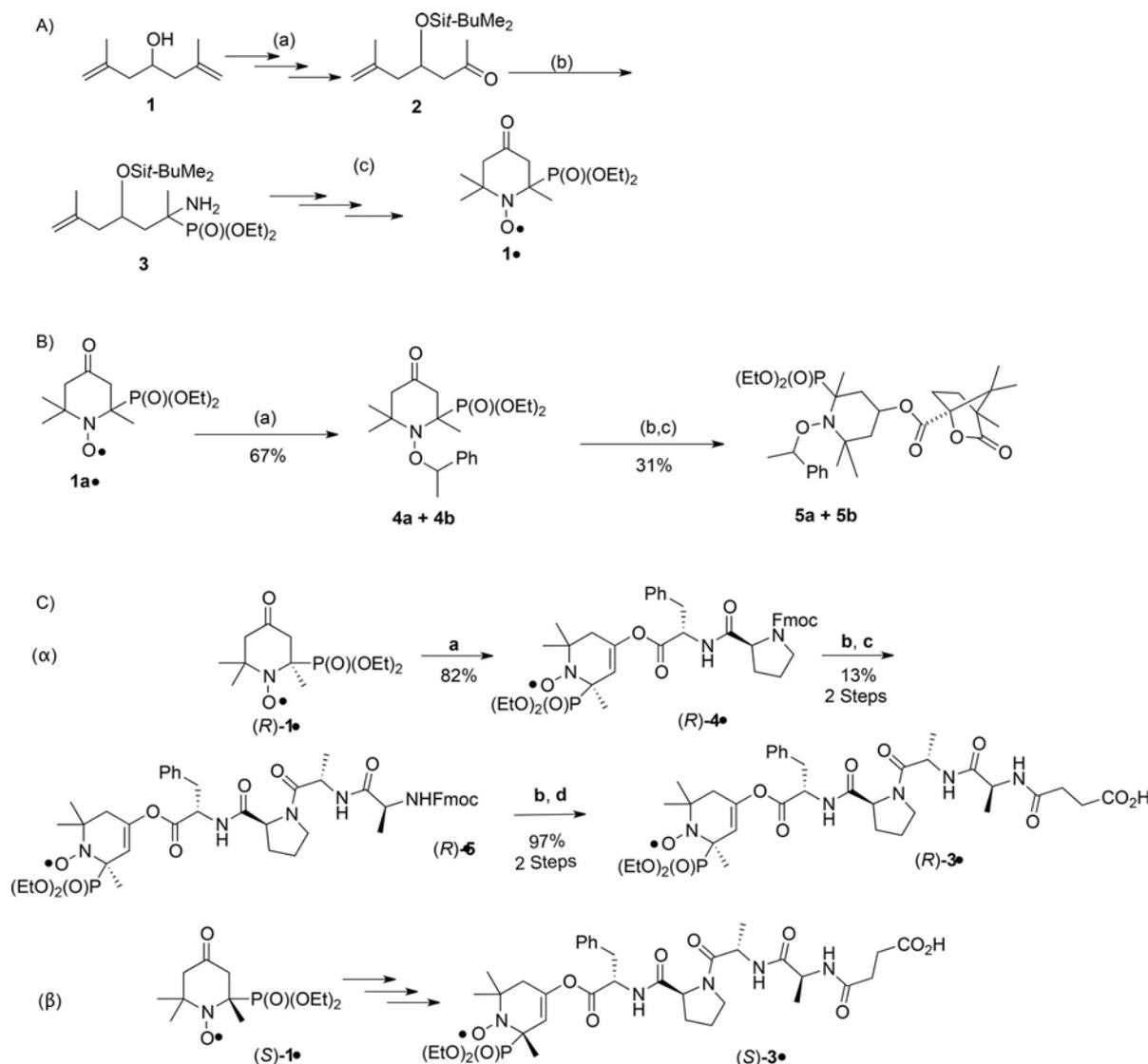


Figure 1. X-ray structures of **5a**^[29] and absolute stereochemistry of **1a**'.

Attempts to prepare (*R*)-**3**' or (*S*)-**3**' using either a step-by-step growth of the peptidic chain or the coupling between the enolate of **1**' and the beforehand-prepared activated tetrapeptide Fmoc-Ala-Ala-Pro-Phe-X failed to yield the aimed compounds. Thus, nitroxide (*R*)-**1**' was treated^[19] with excess of LiHMDS at -78°C and the resulting enolate was trapped with the activated dipeptide Fmoc-L-Pro-L-Phe-O-Piv **6**, prepared by treating the commercially available dipeptide Fmoc-L-Pro-L-Phe-OH with pivaloyl chloride,^[30,31] to yield peptide (*R*)-**4**' (82%). 1,8-Diazabicyclo[5.4.0]undec-7-ene (DBU)^[32] mediated Fmoc deprotection of (*R*)-**4**', followed by *N,N'*-dicyclohexylcarbodiimide (DCC) coupling with commercially available dipeptide, Fmoc-Ala-Ala-OH afforded the tetrapeptide nitroxide (*R*)-**5**' (13% for two steps). Peptide nitroxide (*R*)-**5**' was then treated with DBU, and then by succinic anhydride in a one-pot procedure to yield substrate (*R*)-**3**' as a DBU salt (Scheme 2C α). The same procedure was applied to (*S*)-**1**' to afford (*S*)-**3**' (Scheme 2C β).

EPR signal of substrates (*R*)-**3**' or (*S*)-**3**' display the expected 6-lines signal (top signal in Figure 2) due to the coupling of odd electron with the nitrogen ($I_N=1$) and phosphorous ($I_P=1/2$) atoms (few percent of hydrolyzed nitroxide **1**' are also observed). Nitrogen and phosphorus hyperfine coupling constants a_N ($a_N=15.5\text{ G}$) and a_P ($a_P=38.4\text{ G}$), respectively, are very close to those reported for **2**^[33] as well as the Landé's factor g ($g=2.0057$). Upon addition of chymotrypsin, a new pattern arose (Figure 2) comprising signals of both (*R*)-**3**' and (*R*)-**1**'. After a while, complete hydrolysis of (*R*)-**3**' was observed as only the EPR signal^[19] of **1**' is recorded (bottom signal in Figure 2).

The clear difference between substrate **3**' and the hydrolyzed nitroxide **1**' observed by EPR affords an easy quantification of each species over time. Thus, the selectivity of (*R*)-**3**' and (*S*)-**3**' was investigated with several serine proteases: chymotrypsin, trypsin, porcine pancreatic elastase (PPE), cathepsin G and neutrophil elastase (NE). Initial velocity values show a



Scheme 2. A) a) 1) DMF, imidazole, *tert*-butyldimethylsilyl chloride (TBDMSCl), 5 h, 0 °C; 2) OsO₄, K₃[Fe(CN)₆], K₂CO₃, *t*BuOH/H₂O (1:1), 0 °C, 4.5 h; 3) NaIO₄, THF/water (1:3, v/v), 3 h, 0 °C. b) FeCl₃ (10 mol%), HP(O)(OEt)₂, NH₃, 4 Å MS, 24 h, RT. c) 1) Hg(OAc)₂, THF/water (1:3, v/v), 30 min, RT; 2) NaBH₄, NaOH (1 M); 3) *meta*-chloroperoxybenzoic acid (mCPBA), CH₂Cl₂, 2 h, 0 °C; 4) TBAF, THF, 3 h, 0 °C; 5) NMO, TPAP, 4 Å MS, CH₂Cl₂, 0 °C. B) Determination of the configuration of 1a•. (a) Cu, CuBr₂, PMEDTA, benzene, rt, 12 h. (b) NaBH₄/MeOH, THF, 0 °C, 2 h; (c) (-)-camphanoyl chloride, Et₃N, dry CH₂Cl₂, 0 °C, 1 h. C) Synthesis of chymotrypsin substrate (R)-3• (α) and (S)-3• (β). Reagents and conditions: a) LiHMDS, 6, THF, -78 to -45 °C; b) DBU, DCM, 0 °C, 3 h; c) Fmoc-L-Ala-L-Ala-OH, DCC, HOBT, DCM, 0 °C to RT, 18 h; d) succinic anhydride, DCM, 0 °C to RT, 6 h.

strong selectivity of the substrate (R)-3• for chymotrypsin and cathepsin G (Figure 3). The rates of consumption of (R)-3• by other proteases such as trypsin, PPE and HNE do not differ significantly from its spontaneous hydrolysis (SD in Figure 3). Interestingly, at concentration as low as 2 nM, protease chymotrypsin and neutrophil protease cathepsin G exhibit a high rate of hydrolysis at $V_0 = 24(\pm 2) \text{ nM s}^{-1}$. This selectivity is identical to the one displayed by *para*-nitroanilide (*p*NA) substrates and stems from the almost exclusive preference of these two enzymes for large aromatic amino-acids at the P₁ position.^[34] Similar selectivity is observed for (S)-3•.

For the determination of the Michaelis constants K_M and k_{cat} with chymotrypsin, substrate to product conversion was monitored using EPR spectroscopy by varying the concentration of substrates 3•, at constant concentration of chymotrypsin (Fig-

ure 4a and b). Initial velocities V_0 were measured for each curve and plotted against the concentration of nitroxide (Michaelis-Menten plots Figure 4c). Michaelis constants K_M and catalysis rate constant k_{cat} were estimated by fitting with [Eq. (1)]:

$$V_0 = \frac{k_{cat} \cdot [E][S]_0}{K_M + [S]_0} \quad (1)$$

As shown in Table 1, both enantiomers display comparable values for the Michaelis constant and, hence, a very similar catalytic efficiency k_{cat}/K_M of $3.5(\pm 1.0) \cdot 10^6 \text{ s}^{-1} \text{ M}^{-1}$. It is worth noting that 3• exhibits a 15- to 18-fold higher values for K_M than reported for the widely used chromogenic substrate Suc-Ala-Ala-Pro-Phe-*p*NA along with a 6- to 11-fold higher catalytic

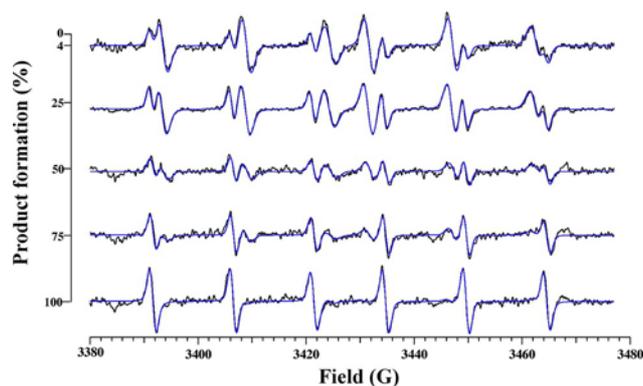


Figure 2. EPR signals of the proteolysis of 50 μM substrate (*R*-3' by chymotrypsin (2 nM) to product 1' (conversion from top to down: 4, 25, 50, 75, 100%). Blue lines are for the EPR simulations using hyperfine coupling constants given in the text and in ref. [19] (see the Supporting Information for details).

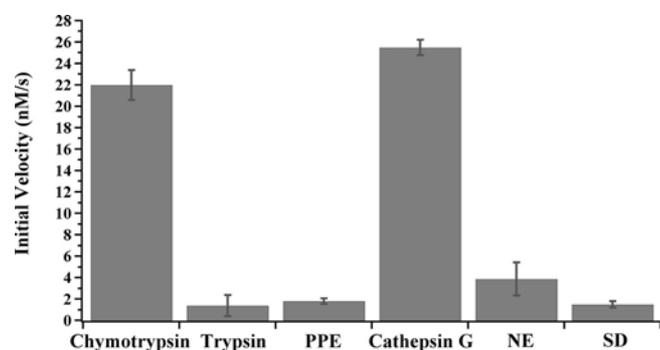


Figure 3. EPR comparative kinetics of hydrolysis of the substrate isomer (*R*-3' (1 mM) by five serine proteases (2 nM) at 25 °C: chymotrypsin, trypsin, porcine pancreatic elastase (PPE), cathepsin G and neutrophil elastase (NE). Substrate spontaneous dissociation in HEPES buffer is given as SD. Error bars for the two limit values of duplicate experiments.

Table 1. Enzyme–substrate Michaelis constants for (<i>R</i>)-3' and (<i>S</i>)-3'.		
	(<i>R</i>)-3'	(<i>S</i>)-3'
K_M [μM]	6 ± 1.6	5 ± 1.2
k_{cat} [s^{-1}]	15 ± 0.7	22 ± 1.1
k_{cat}/K_M [$\text{s}^{-1} \text{M}^{-1}$]	2500 000	4400 000

efficiency.^[2] These significantly better affinity and turnover occurring between chymotrypsin and substrates 3' suggests that the nitroxide leaving group has a stronger interaction with the S' part of the enzyme than the *para*-nitroanilide group according to the Schechter and Berger nomenclature.^[35]

In summary, these experiments exemplify that on/off nitroxides are suitable to probe nonradical enzymatic activity. Moreover, as EPR is able to probe nontransparent media such as blood or tissue samples from biopsies or from medical research experiments, and reports only on radical species (and here only stable free organic radicals) affording a very simple signal irrespective of the conditions. As the substrates reported above show negligible hydrolysis at pH 7.4 they are legible for

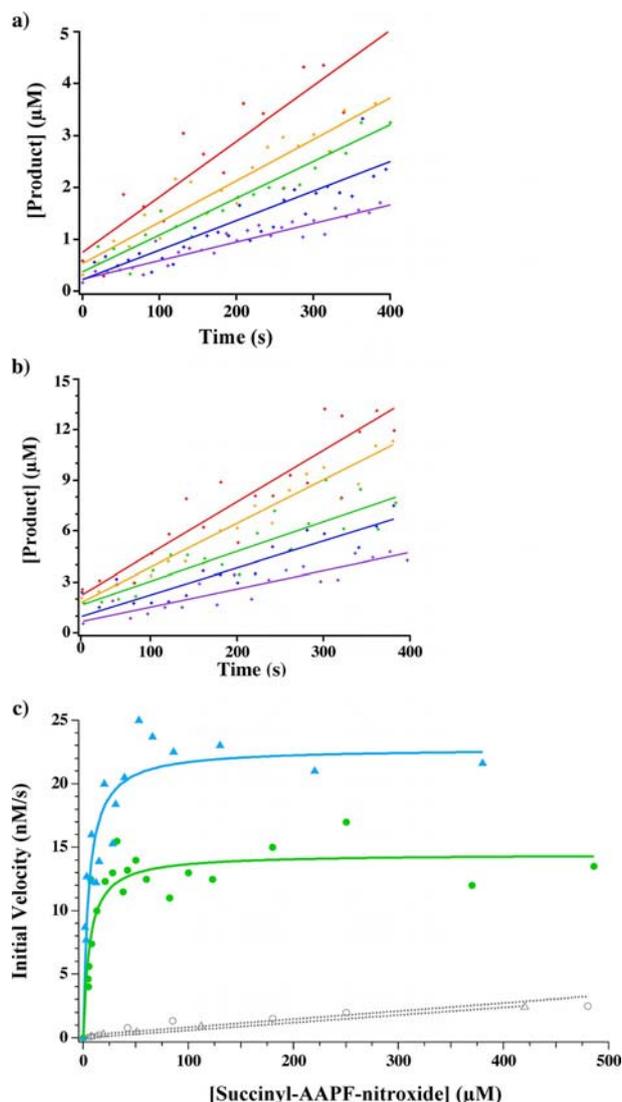


Figure 4. Selected curves of product generation kinetics from a range of substrates 3' concentrations (5 to 250 μM) in the presence of chymotrypsin (2 nM) in HEPES buffer pH 7.4 at 25 °C. a) (*R*-3' (red, orange, green, blue, violet symbols and lines for concentrations of 250, 120, 40, 10, and 5 μM); b) (*S*-3' (red, orange, green, blue, violet symbols and lines for concentrations of 130, 70, 30, 15, and 10 μM). c) Initial velocities (V_0) against substrate 3' concentrations (\blacktriangle for (*S*-3', and \bullet for (*R*-3') in the presence of chymotrypsin (2 nM). Empty symbols for spontaneous hydrolysis. In (a) and (b), full lines are linear fit, and in (c) full lines (green for (*R*-3' and blue for (*S*-3') were fitted with Equation (1).

in vivo and ex vivo experiments. Hence, this method is a convenient approach for the detection of enzymatic activity in samples without any special preparation or denaturation of the samples.

Recently,^[19] we reported on the imaging of enzymatic activity by OMRI using a nonselective probe. Thus, as the high sensitivity to chymotrypsin activity of this substrate is very promising for future imaging of pancreatitis, a pancreatic disease resulting in a premature activation of the pancreatic digestive enzymes inside the pancreas. Imaging of this activation would allow an early diagnosis of the chronic form of the disease. In addition, the selectivity for cathepsin G, a neutrophil protease

which is a reliable inflammation marker, opens the door for a sensitive imaging method of inflammation.

Importantly, the procedure described for the preparation of (S)-3' and (R)-3' is robust enough to be applied to any type of peptides increasing the versatility of our probes provided the nitroxide is stable in the experimental medium. Thus, this approach can be extended to many types of enzymes in various fields of research.

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