Trained Immunity Carried by Non-immune Cells
Attoumani Hamada, Cedric Torre, Michel Drancourt, Eric Ghigo

To cite this version:

Attoumani Hamada, Cedric Torre, Michel Drancourt, Eric Ghigo. Trained Immunity Carried by Non-immune Cells. Frontiers in Microbiology, Frontiers Media, 2019, 9, 10.3389/fmicb.2018.03225. hal-02243365

HAL Id: hal-02243365
https://hal-amu.archives-ouvertes.fr/hal-02243365
Submitted on 21 Nov 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Distributed under a Creative Commons Attribution 4.0 International License
Trained Immunity Carried by Non-immune Cells

Attoumani Hamada, Cédric Torre, Michel Drancourt* and Eric Ghigo

IRD, MEPHI, Institut Hospitalier Universitaire Méditerranée Infection, Aix-Marseille University, Marseille, France

“Trained immunity” is a term proposed by Netea to describe the ability of an organism to develop an exacerbated immunological response to protect against a second infection independent of the adaptative immunity. This immunological memory can last from 1 week to several months and is only described in innate immune cells such as monocytes, macrophages, and natural killer cells. Paradoxically, the lifespan of these cells in the blood is shorter than the duration of trained immunity. This observation suggested that trained immunity could be carried by long lifespan cells such as stem cells and non-immune cells like fibroblasts. It is now evident that in addition to performing their putative function in the development and maintenance of tissue homeostasis, non-immune cells also play an important role in the response to pathogens by producing anti-microbial factors, with long-term inflammation suggesting that non-immune cells can be trained to confer long-lasting immunological memory. This review provides a summary of the current relevant knowledge about the cells which possess immunological memory and discusses the possibility that non-immune cells may carry immunological memory and mechanisms that might be involved.

Keywords: trained immunity, immunomodulatory, non-immune cells, stem cells, lifespan

INTRODUCTION

Works regarding T cell and B cell-independent immune memory date back over half a century (Box 1). A first report by Mackaness (1968) showed that mice vaccinated against tuberculosis with the vaccine bacillus Calmette-Guérin (BCG) were effectively protected against a second infection by mycobacteria. Further, it was shown that trained immunity lasted for weeks to months (Cassone, 2018) and other infectious agents such as Salmonella typhimurium, Listeria monocytogenes, Staphylococcus aureus, Candida albicans, and Schistosoma mansoni (Blanden et al., 1969). These results were confirmed by Tribouley et al. showing the protective effect of BCG on athymic mice against Schistosoma mansoni (Tribouley et al., 1978). In the 80–90s, Bistoni and his colleagues showed that mice infected with attenuated Candida albicans exhibited protection against a lethal dose of Candida albicans and other pathogens such as Staphylococcus aureus (Bistoni et al., 1986). This protection was independent of acquired adaptive immune cells (Box 2) but depended on the innate immune cells as macrophages and a higher production of pro-inflammatory cytokines including interleukin (IL)-1, granulocyte macrophage colony stimulating factor (GM-CSF), tumor necrosis factor (TNF)-α and interferon (IFN)-γ (Bistoni et al., 1986; Vecchiarelli et al., 1988). Then, several studies have shown that in the same way as monocytes, NK-cells exhibit immunological memory. O’Leary et al. showed that a hapten (small molecule triggering an immune response) (Erkes and Selvan, 2014) induced contact hypersensitivity in T and B cell-deficient mice during the second contact with same hapten (O’Leary et al., 2006). This activity was shown to be carried by a liver...
subpopulation of NK cells (Ly49C-I+) (O’Leary et al., 2006). Perforin and granzyme were the factors related to the defense mechanisms of NK-cells (Salcedo et al., 1993). The production of these effectors are controlled by promoter of gene, regulator sequence (enhancer–silencer) and transcription factors such as lymphotoxin α (LTA), tumor necrosis factor α (TNFα), lymphotoxin β (LTB) and interferon-γ (IFNG) genes which may undergo epigenetic modifications (Cichocki et al., 2013; Wiencke et al., 2016). It needs to be investigated how the mechanisms of production of perforin and granzyme may be supported by an epigenetic modification. As for mast cells, there is no published study related to immunological memory. Nevertheless, Monticelli hypothesized that mast cells, like the other innate immune cells, could adopt a memory phenotype (Monticelli and Leoni, 2017). Indeed, mast cells play an important role in the first line defense and possess a longer life than monocyte/macrophage and NK-cells. Also, mast cell biology can be regulated by epigenetic modifications as described in monocytes/macrophages and NK-cells. They play a critical role in the establishment and maintenance of mast cell identity, expansion, differentiation and regulation of mast cell response to a danger signal (Montagner et al., 2016; Monticelli and Leoni, 2017). A recent study reports that the DNMT3A DNA methyltransferase is important to modulate mast cell responses to chronic stimuli (Leoni et al., 2017). Therefore, mast cells could support an immunological memory, which remains to be investigated. At last, dendritic cells (DCs) possess the immunological arsenal against bacteria; have a longer lifespan than monocytes and epigenetic mechanisms to support trained immunity. However, there is no study reporting trained immunity for DCs.

In Kleinnijenhuis et al. (2012) highlighted the mechanism involved in the immune protection previously observed by Bistoni and others. Indeed, these authors showed that reprogramming cells and inflammatory response conferred to monocytes/macrophages were associated with epigenetic modification mechanisms. Briefly, naïve cells have a compact DNA rendering it inaccessible to promoters/ enhancers. After infection, the DNA decondensates making it accessible to the promoters/enhancers that could be mono-methylated and mono-acetylated in a way such that the DNA will be transcribed, allowing for the production of molecules involved in pathogen elimination (Mehta and Jeffrey, 2015). Once the infection is resolved, the acetylation is lost but methylation of lysine 4 of histone 3 (H3K4 me) persists and keeps the promoters active so that during the second stimulation the DNA is rapidly transcribed allowing great production of genes involved in immunity (Ostuni et al., 2013; Quintin et al., 2014; Saeed et al., 2014). Metabolic induction is additional mechanisms underlying trained immunity (Bekkering et al., 2018).

Following pioneering work by Netea, the concept of “trained immunity or innate immune memory” has been proposed (Netea et al., 2011) (Box 3).

Recently, Cheng et al. showed a new mechanism to support “trained immunity” focusing especially on the metabolism of cells such as glycolysis. For example they showed that the cholesterol synthesis pathway was highly induced in β-glucan-treated macrophages (Cheng et al., 2014). The interplay between metabolite production and trained immunity has been recently shown (Arts et al., 2016a). Cholesterol synthesis pathway requires an intermediate metabolite called mevalonate; which induces a
trained immunity profile by promoting the expression of a set of genes required in β-glucan-trained immunity phenotype like fumurate (Arts et al., 2016a; Bekkering et al., 2018). Trained immunity induces an aerobic glycolysis associated with an augmentation of glucose consumption, lactate production and elevated intracellular ratio of nicotinamide adenine dinucleotide (NAD+/NADH) (Cheng et al., 2014).

Trained immunity draws more and more attention and interest. A simple investigation of the number of publications based on the key words “innate immune memory” between 1969 and 2018 shows a sharp increase in the number of papers, reaching 142 papers in July 2018 (Figure 1).

Despite new knowledge, fundamental questions need to be addressed about the paradox between the duration of trained immunity and the short lifespan of innate immune cells (Table 1). Until now all of the studies were performed in monocytes/macrophages and NK cells whereas trained immunity can last for 1 year and more.

Consequently, trained immunity is more and more investigated in non-immune cells such as stem cells, which possess immune characteristics (expression of TLRs, inflammatory response, production of antimicrobial peptides) with long life span.

**Epigenetic Mechanism**

Term composed by “epi” meaning “above” in Greek and “genetic” relating to genes. Basically, it is the set of chemical modifications occurring in the DNA and consequently modulating the expression of genes (Box 4). The mechanism does not affect the sequence of the DNA but is transmissible to the offspring. Epigenetic modifications include DNA methylation, histone methylation and acetylation (Saeed et al., 2014; Hoeksema and de Winther, 2016). In general, DNA methylation is an epigenetic mechanism which involves the addition of a methyl-CH3 group on carbon predominantly to the CpG dinucleotides of the cytosine residues of DNA 5-methylcytosine (5 mC). This process involving three DNA methyltransferases (DNMT1, DNMT3A, and DNMT3B) is active in the regulation and maintenance of gene expression (Jaenisch and Bird, 2003). NK-cell memory trained by BCG is associated with DNA methylation (Sun et al., 2012; Schlums et al., 2015).

**Immunometabolic Mechanism**

Contraction of “immune” and “metabolic,” originally proposed to explain both the cellular metabolism of innate immune cells and the role of immune cells playing in metabolic diseases and/or organ metabolism in global. The first study on immunometabolic mechanism concerned the relation between immunity and metabolic diseases, diabetes, and obesity (Ferrante, 2013). Recent studies showed that metabolic mechanisms are also highlighted in trained immunity (Arts et al., 2016b; Hotamisligil, 2017). In monocytes trained by β-glucan, transcriptional and epigenetic analysis revealed an increase in the promoters of genes encoding enzymes involved in glycolysis (hexokinase and pyruvate kinase) and its master regulator mTOR (mammalian target of rapamycin) (Cheng et al., 2014).

**TRAINED IMMUNITY OF NON-IMMUNE CELLS**

**Stem Cells**

**Mesenchymal Stromal/Stem Cells**

Mesenchymal stem cells possess immune properties such as immunosuppressive phenotype, inflammatory phenotype, anti-bacterial characteristics and are equipped with Pattern Recognition Receptors (PRRs), including Toll-Like Receptors (TLRs) (Pevsner-Fischer et al., 2007; Liotta et al., 2008; Coffelt et al., 2009; Krasnodembskaya et al., 2010; Machado et al., 2013). In 2006, Guang-yang Liu and his colleagues showed that adipose mesenchymal stem cells exhibit short-term memory when exposed a second time to bacterial ligand LPS or danger signal TNF-α. Precisely, mesenchymal stem cells primed with LPS or TNF have the ability to produce more intensely pro-inflammatory cytokines IL-8, MCP-1, and IL-6 to the same stimulus upon a second encounter. Additionally, they showed that primed mesenchymal stem cells exhibited a better therapeutic effect on diabetic rat model than unprimed MSCs (Liu et al., 2016). Moreover, like in monocytes, this trained immunity is governed by epigenetic mechanisms. The authors showed the involvement of a set of microRNAs (miR146a, miR150, and miR155) and a modification of by 5-hydroxymethylcytosine (5 hmC) (Liu et al., 2016). This study was the first one to demonstrate that trained immunity could be carried by non-professional immune cells. Nevertheless, it concerns a short-term memory only corresponding just to 7 days whereas the trained immunity can last from 1 week to months and several years (Nguipdop-Djomo et al., 2016). Recently, it has been shown that the planarian *Schmidtea mediterranea* is exhibiting trained immunity function (Torre et al., 2017). Planarian previously infected with *Staphylococcus aureus* developed an improved immune response with increased bacterial clearance during a second infection. This immunological memory is due to stem cells that support cell replacement during homeostasis and regeneration of any missing tissue (Eisenhoffer et al., 2008; Kaufmann et al., 2018). RNAi-based experiments revealed the signaling...
<table>
<thead>
<tr>
<th>Types of cells</th>
<th>Life Span</th>
<th>Priming agents</th>
<th>Mechanism underlying</th>
<th>Consequences</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Innate immune cells</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Monocytes Macrophages</td>
<td>7 days</td>
<td>BCG</td>
<td>Epigenetics modifications: (H3K4me3 persistency), Increased H3K4 trimethylation monocytes after BCG Vaccination and training BCG vaccination dependent to NOD2 and Rip2</td>
<td>BCG induces immunological memory protection through reprogramming cells, inflammatory response, increase of cytokine production (IFN-γ, TNF, and IL-1β)</td>
<td>Kleinnijenhuis et al., 2012</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Candida albicans</td>
<td>Epigenetic modifications: increase of H3K4me3.</td>
<td>Protection against reinfection induced by Candida albicans. Pro-inflammatory protective response TNF-alpha, IL-6, and IL-18.</td>
<td>Quintin et al., 2012</td>
</tr>
<tr>
<td>NK-cells</td>
<td>6 Months</td>
<td>Hapten-induced contact</td>
<td>Trained immunity carried by NK-cells (Ly49)</td>
<td>Inflammatory memory induced against hapten 2,4-dinitro-1-fluorobenzene (DNFB) or oxazolone</td>
<td>O’Leary et al., 2006</td>
</tr>
<tr>
<td></td>
<td></td>
<td>MCMV</td>
<td>Reprogramming NK-cell with pro-inflammatory cytokines signals operating through IL-12 and STAT4</td>
<td>MCMV-specific NK cell clonal expansion as well as memory NK cell formation: protection against MCMV infection</td>
<td>Sun et al., 2012</td>
</tr>
<tr>
<td>Non-immune cells</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hematopoietic Cells</td>
<td>Indefinite lifespan</td>
<td>BCG</td>
<td>BCG induce epigenetic modification for three histone marks (H3K4me1, H3K4me3, H3K27AC).</td>
<td>Expansion of HSC, Myelopoiesis, BCG train HSCs to generate trained monocytes/macrophages, high production of cytokine essential for protective antmycobacterial</td>
<td>Kaufmann et al., 2018</td>
</tr>
<tr>
<td></td>
<td></td>
<td>β-glucan</td>
<td>Immunometabolic pathways β-glucan induce an increase of glycolysis in train HSCs.</td>
<td>Expansion of HSPCs, IL-1b GM-CSF</td>
<td>Mitroulis et al., 2018</td>
</tr>
<tr>
<td>Mesenchymal Stem Cells</td>
<td>Indefinite lifespan</td>
<td>LPS</td>
<td>Epigenetic mechanism: miRNAs (miR146a, miR150, and miR155, along with the modification of DNA by 5-hydroxymethylcytosine (5hmC))</td>
<td>Increased expression of pro-inflammatory cytokine IL-6, IL-8</td>
<td>Liu et al., 2018</td>
</tr>
<tr>
<td>Epithelial Stem Cells (EpSCs)</td>
<td>Indefinite lifespan</td>
<td>Imiquimod (IMQ)-induced model of skin inflammation</td>
<td>Epigenetic modifications: induced epithelial stem cells maintains chromosomal accessibility of both epidermal and inflammation genes after the first stimulus, In the second stimulus genes were transcribed rapidly.</td>
<td>Inflammatory memory carried by non-immune cell (EpSCs) of the skin. Accelerating wound repair in induced mice 2.5 times faster than naive.</td>
<td>Naik et al., 2017</td>
</tr>
</tbody>
</table>
Smed-PGRP-2/Smed-setd8-1 methyltransferases as key factors in instructed neoblasts (planarian stem cells) (Torre et al., 2017). This study provided additional information to explain the capacity of BCG to induce a long immune memory (30 days minimum) and to persist for months to years whereas the lifespan of many innate immune cells in circulation is limited on the order of hours or days (Sun et al., 2011; Kleinnijenhuis et al., 2012; Nayar et al., 2015). Eventually, Kaufman et al. showed that priming HSCs with intravenous BCG vaccine reaching the bone marrow induced a protective memory against bacterial pathogen; whereas sub-cutaneous vaccination did not prime HSCs. Priming innate immune progenitor cells promoted myelopoiesis and the generation of trained monocytes/macrophages enhancing bacterial clearance. This was the first study exploring the mechanism of long term effects of BCG on trained immunity (Kaufmann et al., 2018).

Therefore, it is important to study the immunological memory capacity of stem cells beyond 1 month and their ability to transmit information to innate immune cells.

**Hematopoietic Stem Cells**

Kaufmann and her colleagues published an important study which brings additional information to explain how cells transmit their immunological memory to progenitors, providing long-lasting protection (Kaufmann et al., 2018). These authors showed that in mice, the inoculation of bone marrow with BCG induced protection against *Mycobacterium tuberculosis* during a second infection which was not the case after subcutaneous inoculation. Specifically, it was shown that bone marrow BCG inoculation initially increased the amount hematopoietic stem cells (HSCs) and myelopoiesis and trained HSCs which offspring had the ability to develop a memory response against virulent *Mycobacterium tuberculosis*. Moreover, BCG induces epigenetic modifications of 2,483 genes in the macrophages, inducing changes of histone H3K27ac in (BMDM) Bone-marrow-derived macrophage produced from BCG-vaccinated mice compared with PBS-control mice (Kaufmann et al., 2018).

Another recent study showed that trained immunity performed at the level of bone marrow precisely increased myelopoiesis (Mitroulis et al., 2018). A total of 1,383 differentially expressed genes in β-glucan-injected mice compared to PBS-treated control mice, were involved in some innate immune functions such as production of pro-inflammatory cytokine IL-1β, production of GM-CSF (granulocyte-macrophage colony-stimulating factor) and immunometabolism including the biosynthesis of cholesterol and glycolysis (Mitroulis et al., 2018).

**Epithelial Stem Cells**

Epithelial stem cells are progenitors of differentiated epithelial cells. Epithelial cells and fibroblasts are forming a physical and functional barrier against external agents (Sacco et al., 2004). They constitute the first line of defense in innate immunity (Ochiel et al., 2008). Like innate immune cell, epithelial cells can act as sentinel cells by expressing TLRs, producing immunomodulator factors and antimicrobial peptides when exposed to danger signals like bacteria, virus, and noxious signals (Schaefver et al., 2004; Bautista-Hernández et al., 2017). In addition to these immune characteristics, it was reported that skin epithelial stem cells exhibit trained immunity. By using an imiquimod (IMQ)-induced model of skin inflammation, the authors showed that the skin previously exposed to one inflammatory challenge responded faster to an unrelated secondary challenge, with faster wound healing in primed mice than in naïve mice (Naik et al., 2017). A recent review by Novakovic entitled “I Remember You: Epigenetic Priming in Epithelial Stem Cells” speculated that this memory of inflammation could be due to epigenetic mechanisms (Novakovic and Stunnenberg, 2017). Additionally, epithelial cell possess a long lifespan averaging 2 years (Tunn et al., 1989).

**Intestinal Stromal Cells**

Intestinal stromal cells (iSCs) are part of non-professional immune cells including mesenchymal stem cells, fibroblasts, epithelial cells and endothelial cells that exhibit immune properties and contribute to immunity processing. ISC express TLRs 1–9, act as sentinel cells and produce pro-inflammatory cytokines in response to a pathogen (Owens and Simmons, 2013; Augenlicht et al., 2016). Recently, Owens assessed that iSCs possess the immune machinery to respond against bacterial pathogens and suggested that iSCs could exhibit an immunological memory (Owens, 2015). Indeed iSCs could develop a protective response to rapidly eliminate the pathogen or another microorganism during a second contact (Owens, 2015). ISCs produce prolonged pro-inflammatory cytokines (inflammatory memory) to recruit immune cells to the site of infection during a second contact (Owens and Simmons, 2013; Owens et al., 2013; Owens, 2015). Epigenetic modifications and immunometabolic mechanisms would be implicated in the persistence of immunological memory. Moreover, the lifespan of iSCs is longer than professional innate immune cells (Arts et al., 2016b; Hotamisligil, 2017; Smith et al., 2017).

**Fibroblasts**

The main function of fibroblasts is to maintain the structural integrity of the connective tissue (Wong et al., 2007). Nevertheless, it has now been well established that fibroblasts play an important role in the immunity. Fibroblasts are equipped with 1 to 10 TLRs, produce pro-inflammatory and antimicrobial peptides, cytokines, chemokines and growth factors in response to pathogen invasion (Jordana et al., 1994; Van Linthout et al., 2014). Further, studies reported that fibroblasts are sentinel cells responding to pathogens and interacting with other cells via the production of molecular signals (Kaufman et al., 2001). Recent studies revealed that fibroblasts were involved in the persistence of inflammation. When activated with bacterial infection or
danger signal (cytokines), tissue repair triggers a protective immune response by producing cytokines and interacting with immune cells (Flavell et al., 2008; Frank-Bertoncelj et al., 2017). The persistence of inflammation is controlled by epigenetic mechanisms. In response to injury or infection, fibroblasts first recruit immune cells in the site of infection to clear bacteria, then organize tissue repair and renewal (Flavell et al., 2008). Indeed, tissue repair and renewal are not random as fibroblasts possess a positional memory capacity enabling regenerating cells to recall spatial information from the uninjured tissue (Bustos-Arriaga et al., 2011; Van Linthout et al., 2014). This process is governed by epigenetic regulation of the homeobox “HOX” genes also involved in regulating body formation during development (Francis and Kingston, 2001; Coleman and Struhl, 2017). Much remains to be investigated in the immune functions of the fibroblasts such as the immunological memory. Functionally fibroblasts possess the immunological arsenal as mentioned above: the ability to produce inflammatory cytokines, antimicrobial peptides, and possess a longer lifespan than the cells of innate immunity such as macrophage once differentiated (Weissman-Shomer and Fry, 1975).

**Microglial Cells**

Microglial cells are resident immune cells of the central nervous system (CNS) also known as resident macrophages of CNS (Wake and Fields, 2011). The main function of microglial cells is to ensure the homeostasis of synapses and the communication with the micro-environment in the CNS. Once activated by virus or bacteria, microglial cells somal size increases while retraction and thickening of processes are facilitating their migration capacity; they express TLRs, produce inflammatory cytokines and acquire phagocytic ability like macrophages (Mariani and Kielian, 2009). After this priming, microglial cells become susceptible to a secondary danger signal leading an improve immune response (Perry and Holmes, 2014; Haley et al., 2017). Several reviews suggested a process of trained immunity in microglial cells (Haley et al., 2017; Lelios and Greter, 2018; Wendeln et al., 2018). Haley recently speculated the fact that microglial cells possess an innate immune memory characterized by inflammatory pathways orchestrated with epigenetic mechanisms (Haley et al., 2017). A study comparing naive mice and mice primed with attenuated *Salmonella typhimurium* containing its LPS indicated that the second group exhibited increased microglial immunoreactivity in response to a second stimulation of LPS four weeks later. Interestingly, there was no increased immunoreactivity in microglial for the naive mice (Püntener et al., 2012). However, the authors did not investigate a potential epigenetic mechanism to explain the long-term activation of microglia in this infectious model.

**CONCLUSION**

The established dogma that innate immunity system lacks memory and that only the adaptive immune system is able to recognize an infectious agent and to destroy it faster in a second encounter has been challenged. Current published data indicate that innate immune cells are able to build an immunological memory. Indeed, the new concept called “trained immunity” represents a paradigm change in the biology of immunity, emerging as a third way between the conventional dichotomy “innate immunity” and “adaptive immunity.” First report of trained immunity described an inflammatory protection of organism against a pathogenic microorganism upon a second encounter independent of adaptive immunity. It is initially concerned monocytes/macrophages and NK cells. However, in the last decade, it has been demonstrated that stem cells and other stromal cells display immune abilities, limited compared to professional innate immune cells, yet contributing to protective immune responses by humoral mediation (inflammatory). Eventually, trained immunity is also described in non-immune cells such hematopoetic stem cells and mesenchymal stem cells. The common mechanism of the non-immune cells in term of trained immunity is based in the ability to respond inflammatory stimuli. Like professional immune cells, a second challenge activates faster the production of cytokines. Consequently, this inflammatory ameliorates the immune response by rapid activation and recruitment of the immune cells at the site of infection, facilitating wound repair. Quantitative real time qRT-PCR array revealed that epigenetic modification mechanisms are involved in the instruction and establishment of inflammatory memory.

Finally, in order to avoid any confusion and to harmonize the concept of immunological memory, we propose to keep the term “trained immunity” because an increasing number of studies are showing that non-immune cells possess immunological memory which is therefore not restricted to innate immune cells.

**AUTHOR CONTRIBUTIONS**

AH and CT drafted the first version. EG and MD conceived the research and approved the final version.

**ACKNOWLEDGMENTS**

This study was supported by IHU Méditerranée Infection, Marseille, France and by the French Government under the “Investissements d’avenir” (Investments for the Future) program managed by the Agence Nationale de la Recherche (ANR, fr: National Agency for Research), (reference: Méditerranée Infection 10-IAHU-03). This work was supported by Région Provence Alpes Côte d’Azur and European funding FEDER PRIMI. The authors acknowledge Olga Cusack for her expert assistance in editing the manuscript. AH benefits from a PhD grant from Direction des Politiques Scolaires et Universitaires (DPSU), Conseil départemental de Mayotte.